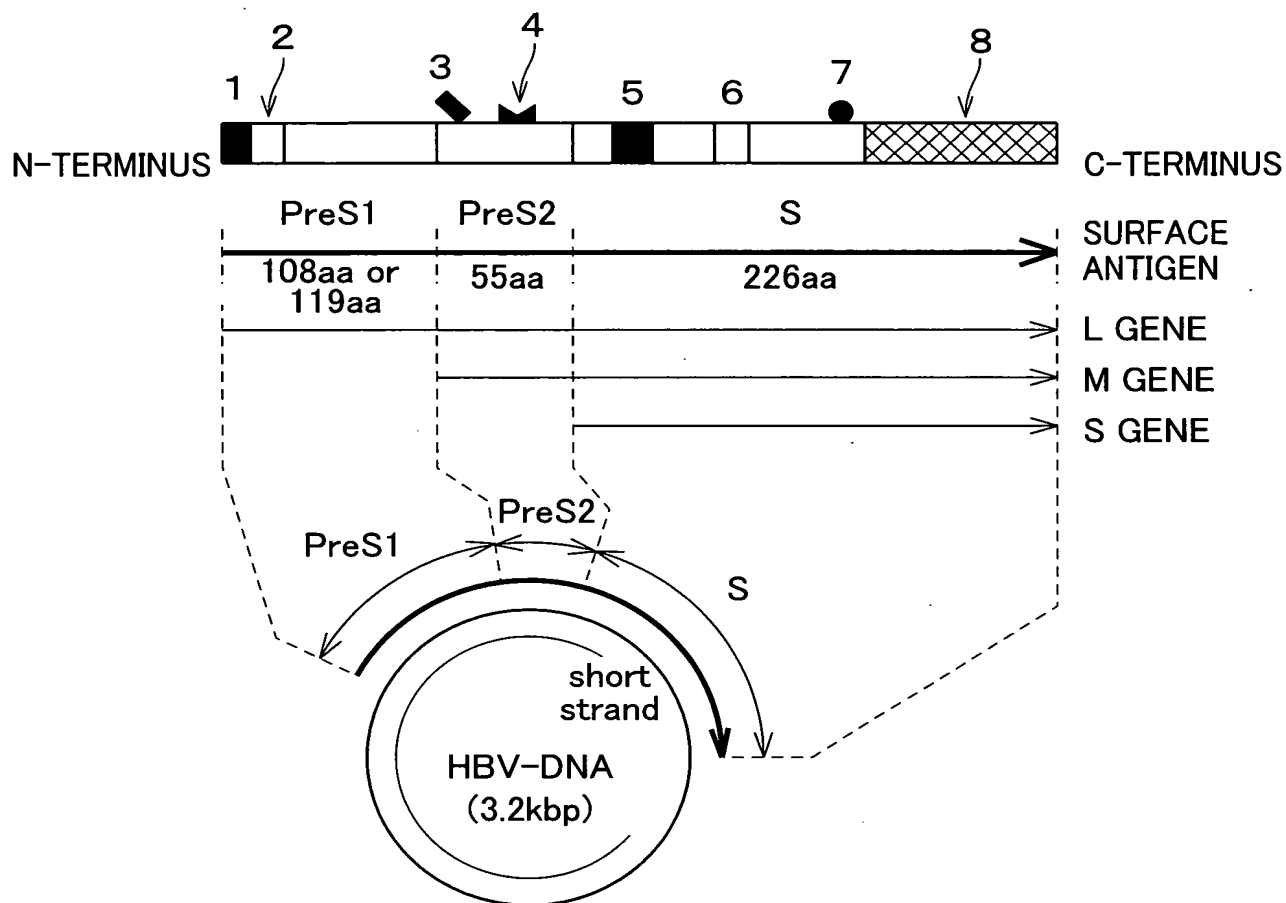


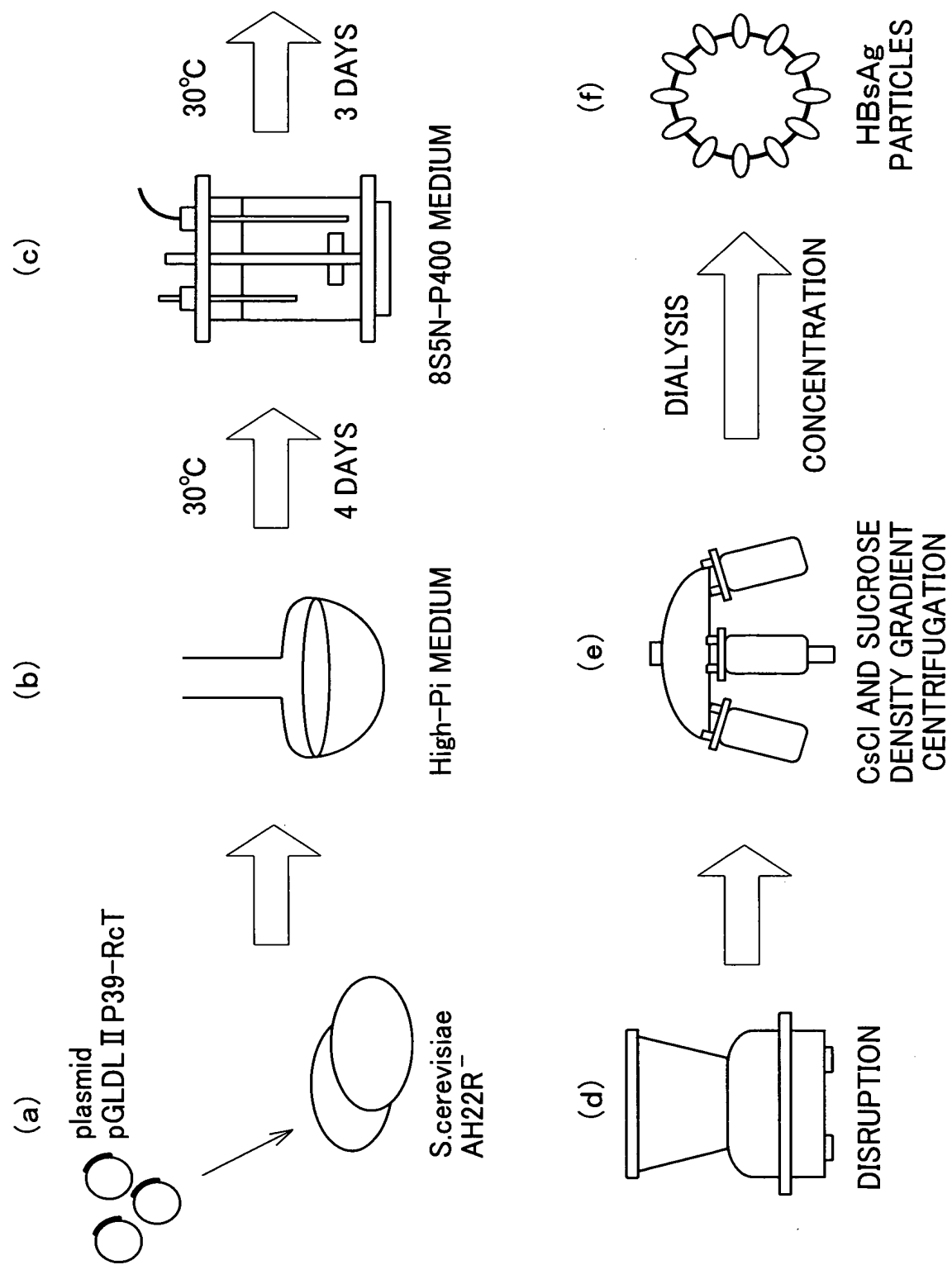
FIG. 1



- 1 PARTICLE FORMATION SUPPRESSING SITE
- 2 DIRECT RECEPTOR SPECIFIC TO HUMAN HEPATOCYTE
- 3 SUGAR CHAIN 1
- 4 INDIRECT HUMAN HEPATOCYTE SPECIFIC RECEPTOR
(POLYMERIZED HUMAN SERUM ALBUMIN RECEPTOR)
- 5 TRANSMEMBRANE REGION 1
- 6 TRANSMEMBRANE REGION 2
- 7 SUGAR CHAIN 2
- 8 TRANSMEMBRANE REGION 3

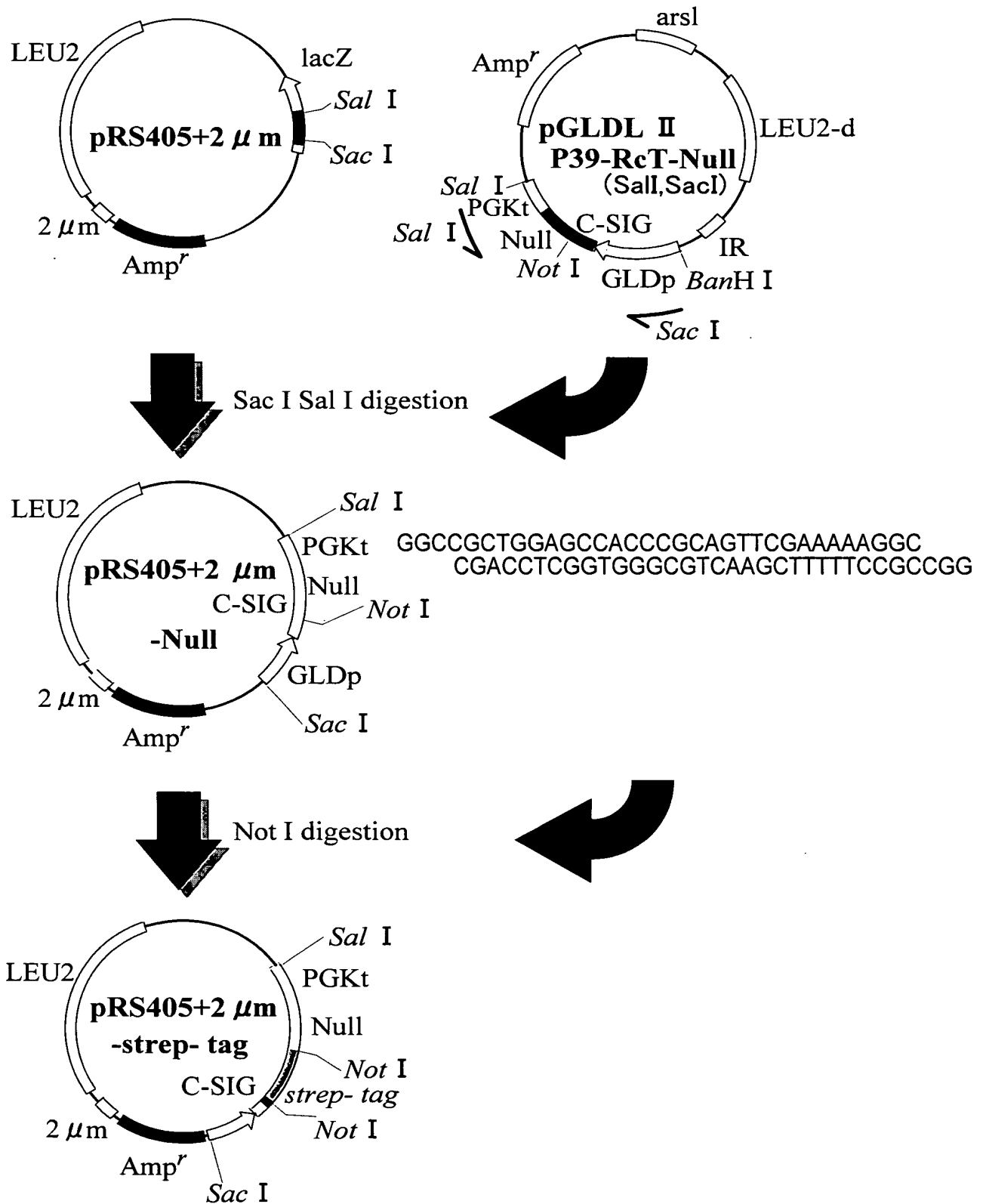
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FIG. 2



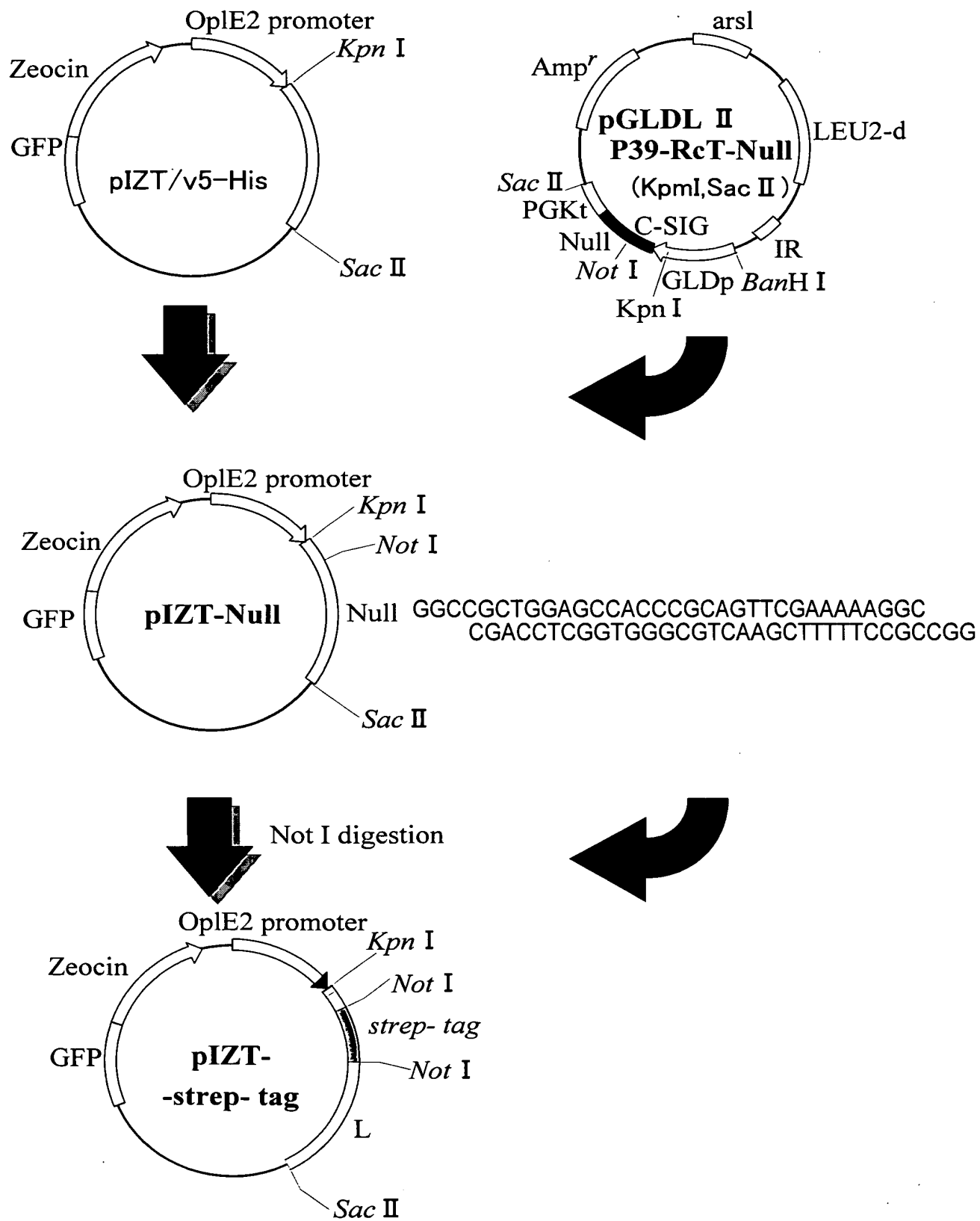
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FIG. 3



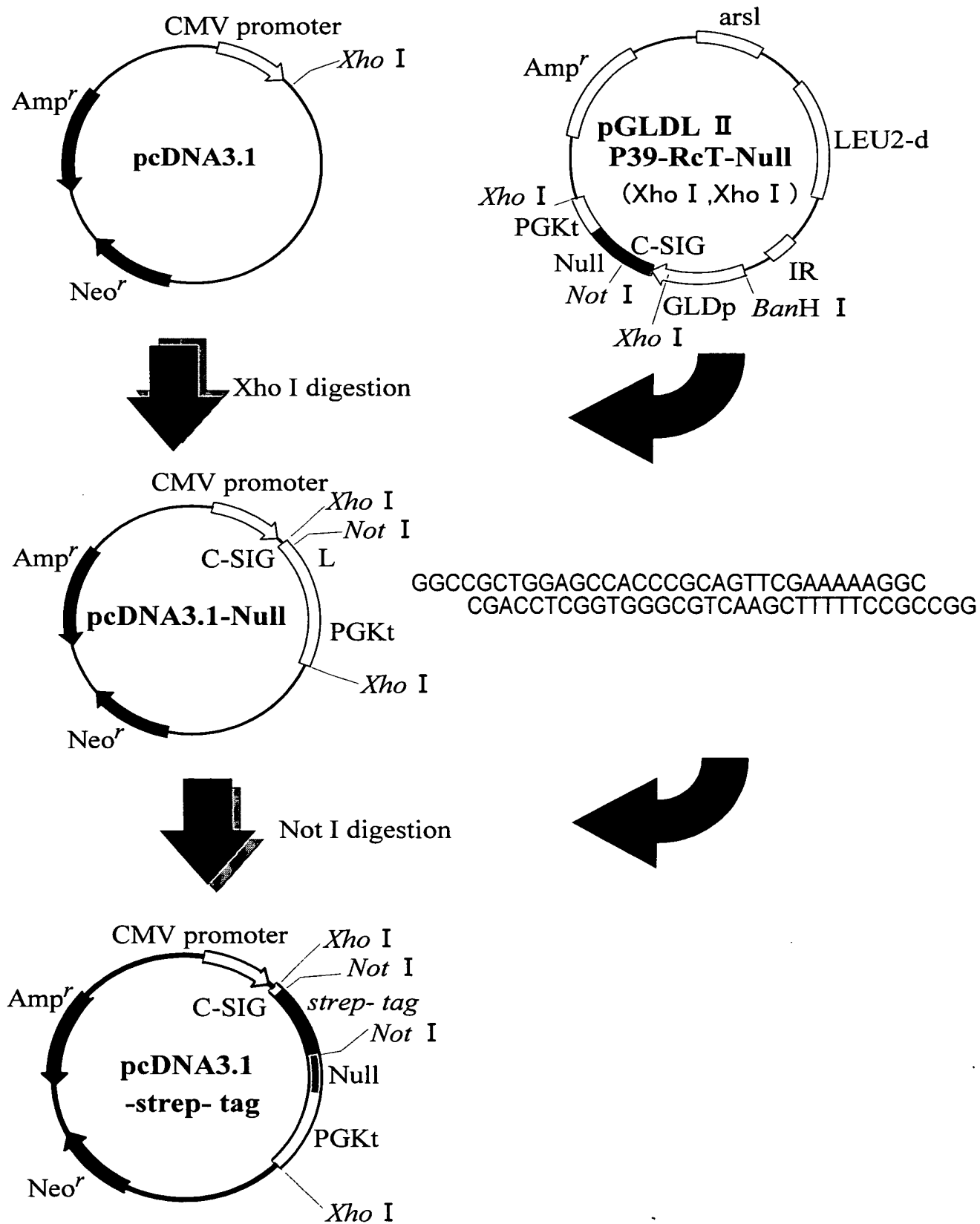
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FIG. 4



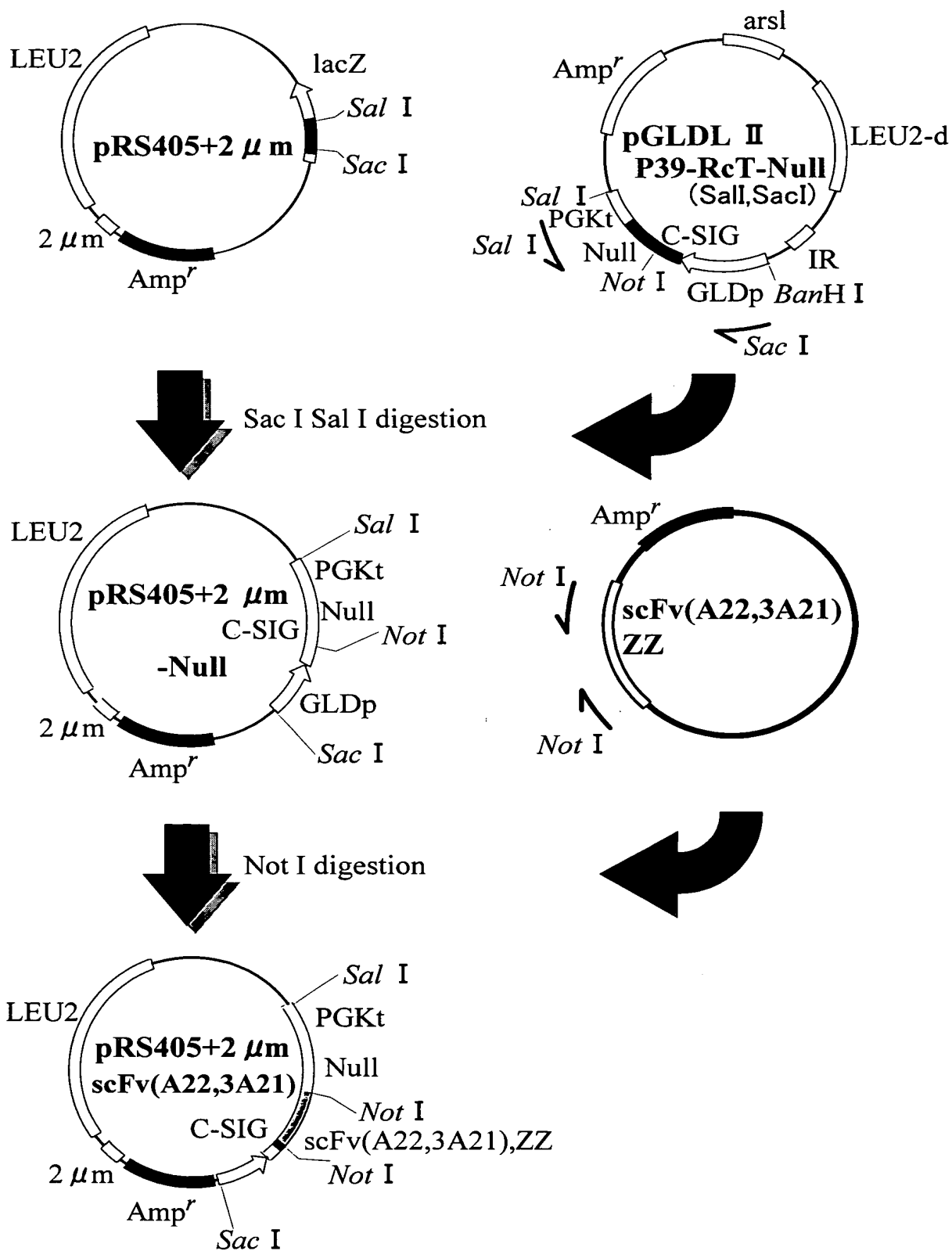
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FIG. 5



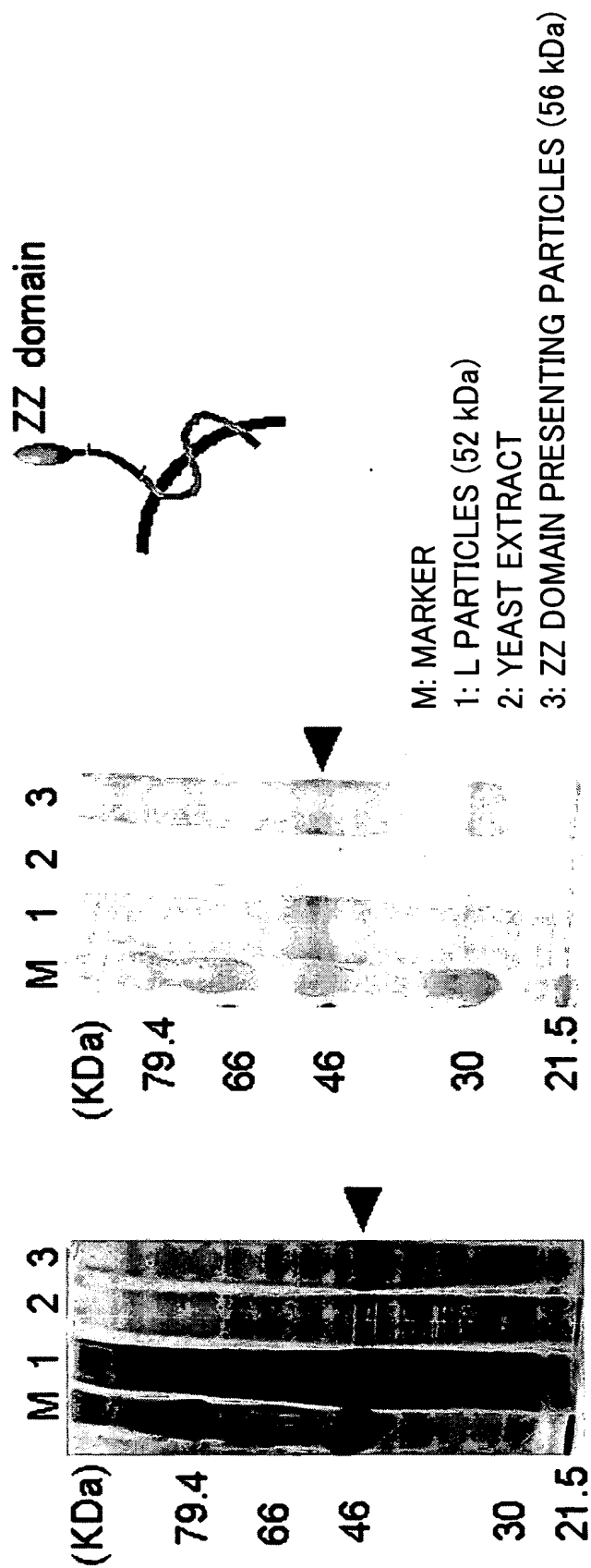
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FIG. 6



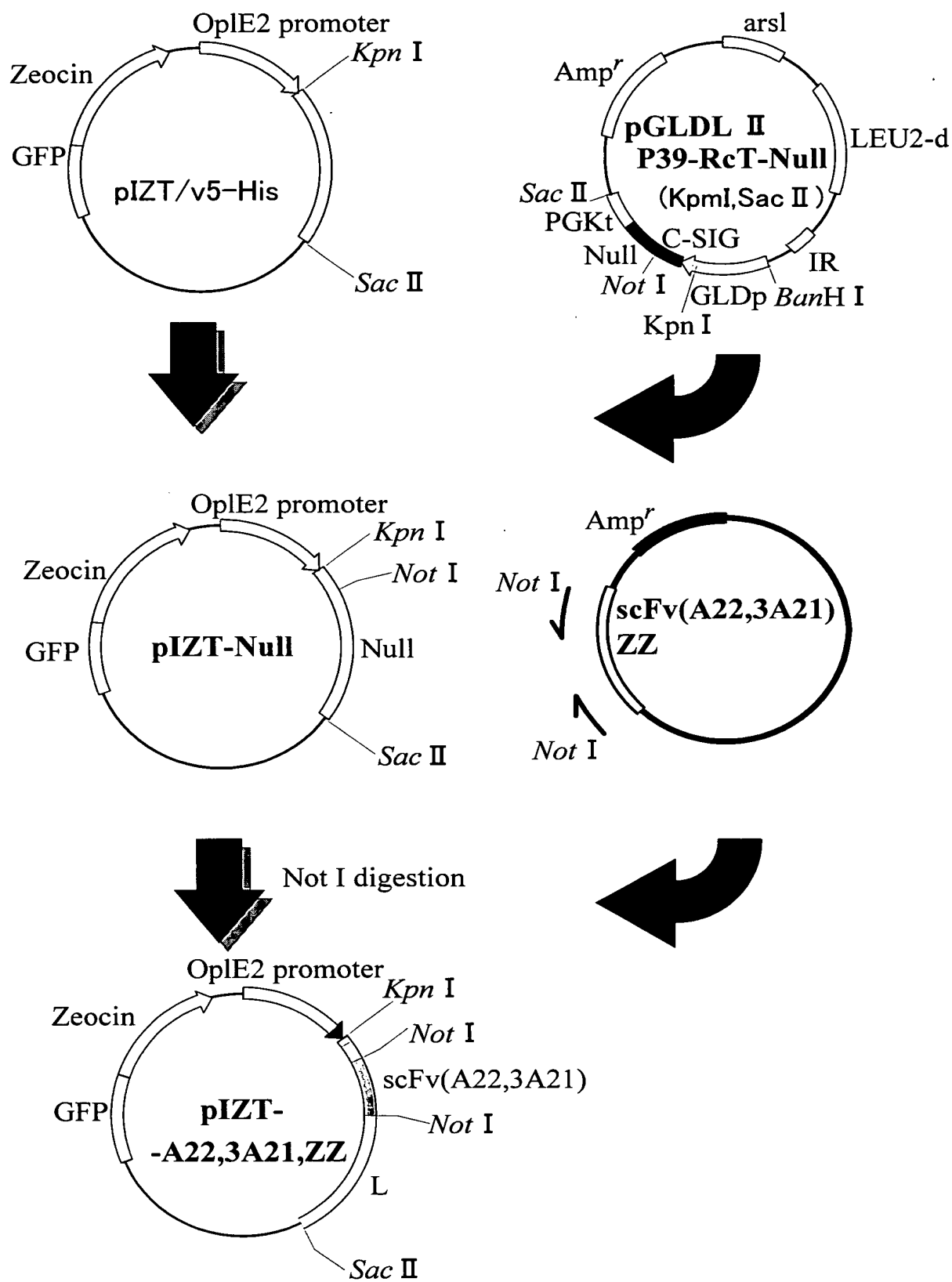
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FIG. 7



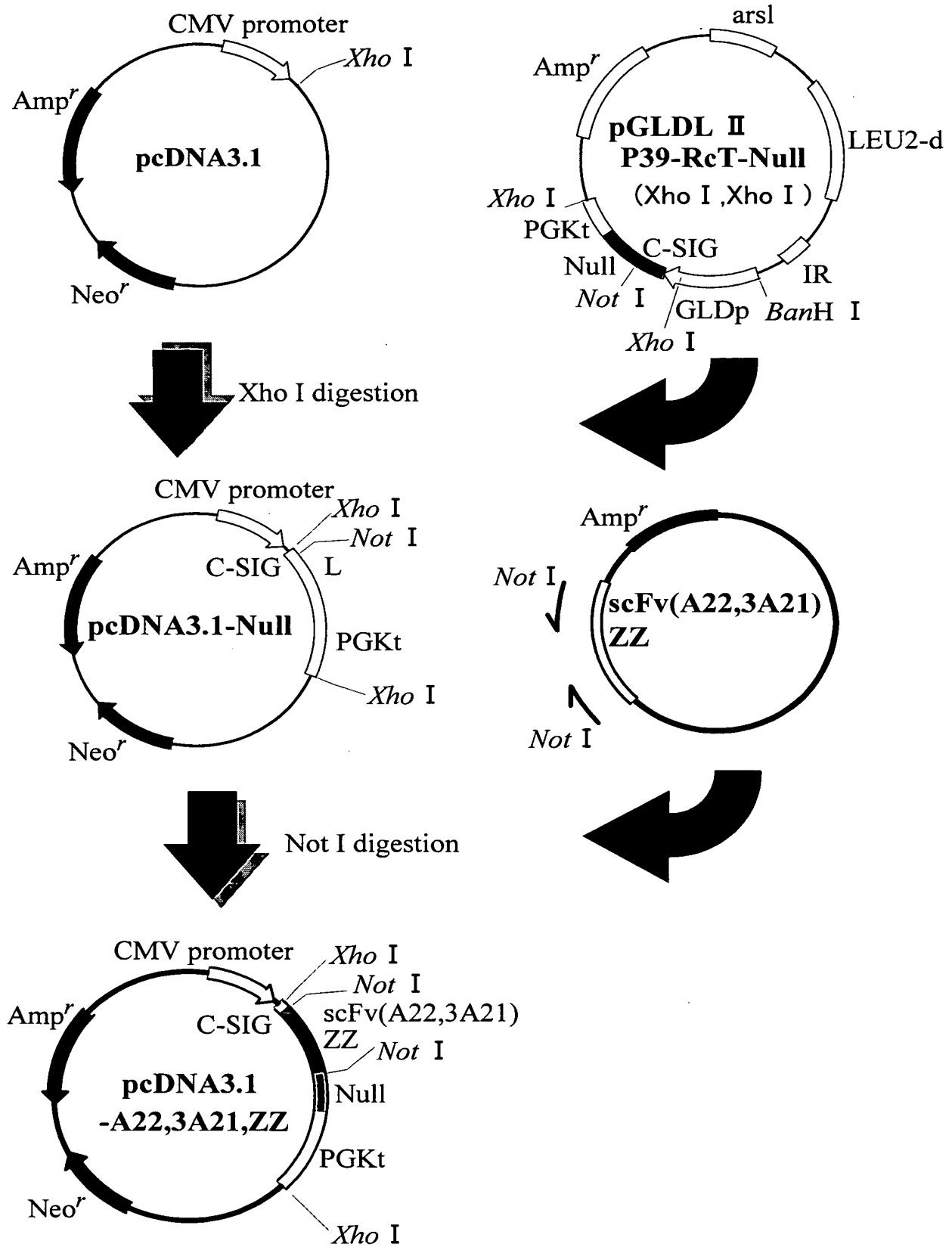
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FIG. 8



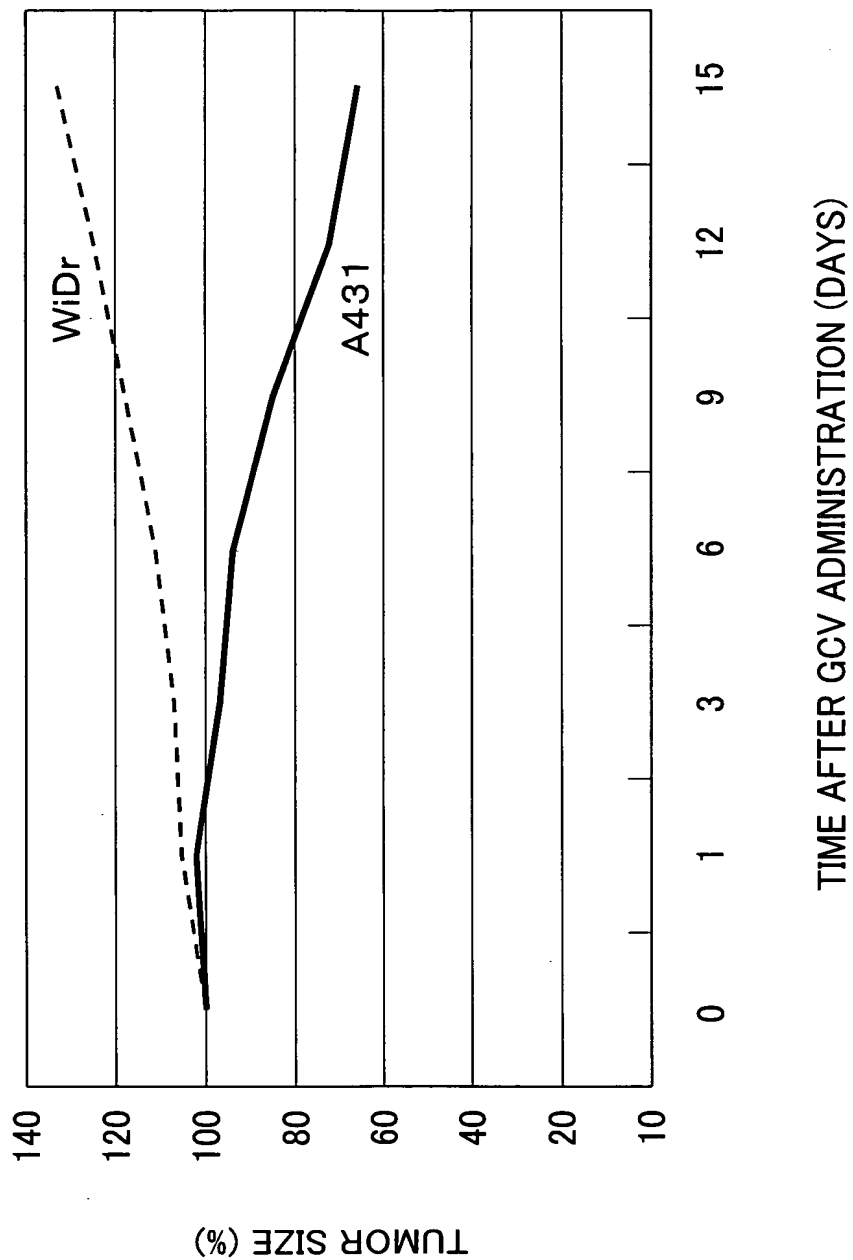
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FIG. 9



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FIG. 10



scFv(A22)

M: MARKER
1: A22 PRESENTING PARTICLES (76 kDa)

(KDa)

121
95
49
40
20
30

M 1

(KDa)

97
66
45
30
20
14

M 1

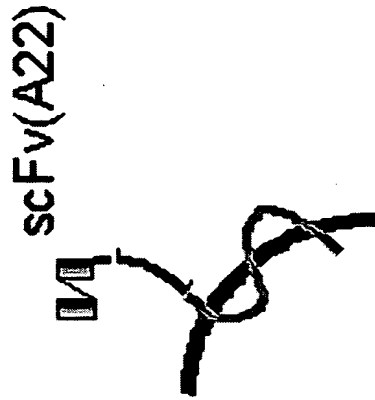


FIG. 12

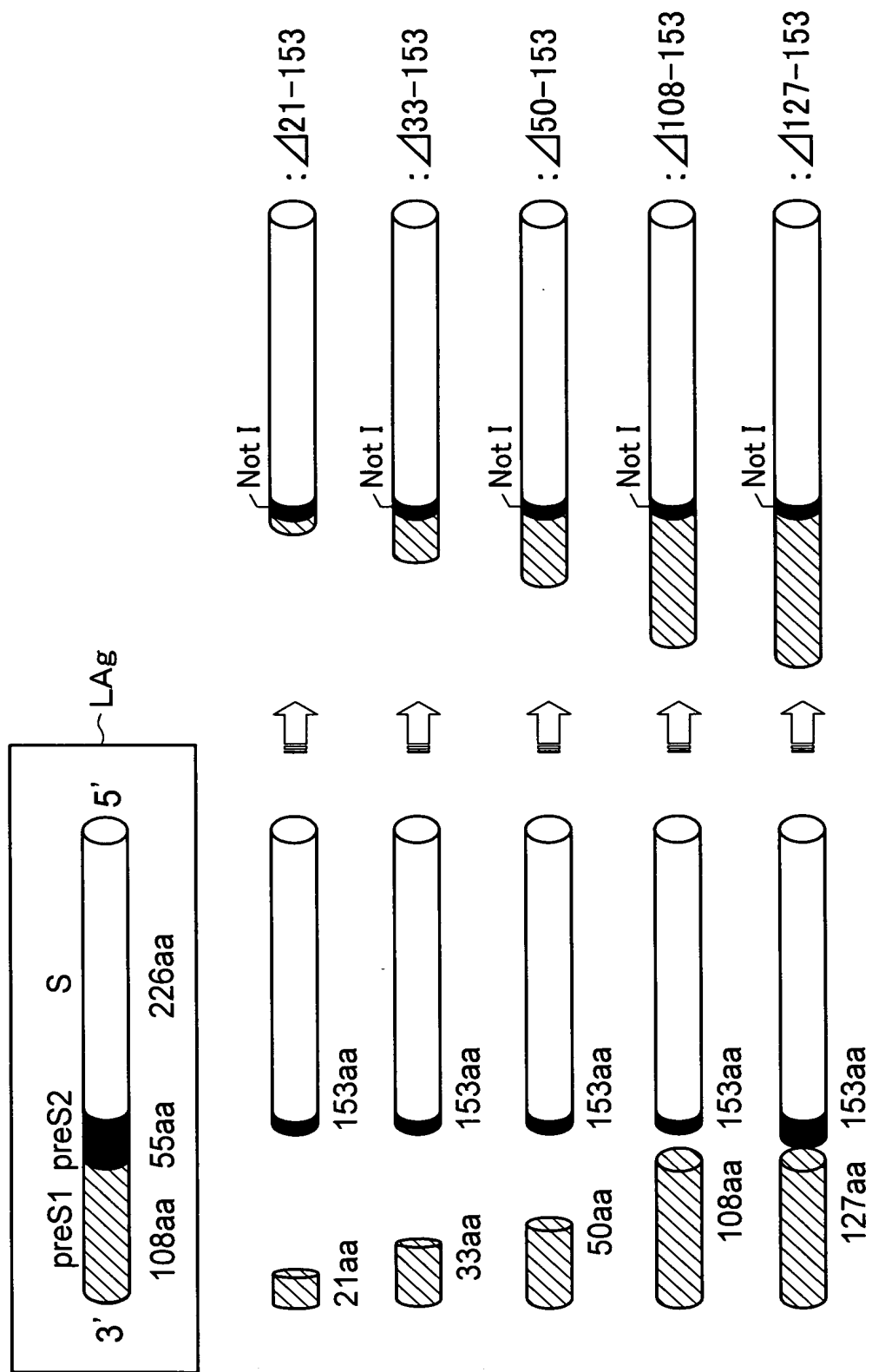


FIG. 13

Pyrobest	DNA	0.5 μ l
POLYMERASE (TaKaRa)		
10 \times PCR BUFFER		5 μ l
dNTP MIXTURE (10mM)		5 μ l
TEMPLATE DNA(5 μ g/ml)		2 μ l
PRIMER (F)(100 μ M)		1 μ l
PRIMER (R)(100 μ M)		1 μ l
DISTILLED WATER		35.5 μ l
TOTAL		50 μ l

FIG. 14

	CYCLE	TEMPERATURE	TIME
1	1	98°C	30 sec.
2	30	98°C	30 sec.
		55°C	1 min.
		68°C	30 min.
3	1	4°C	∞

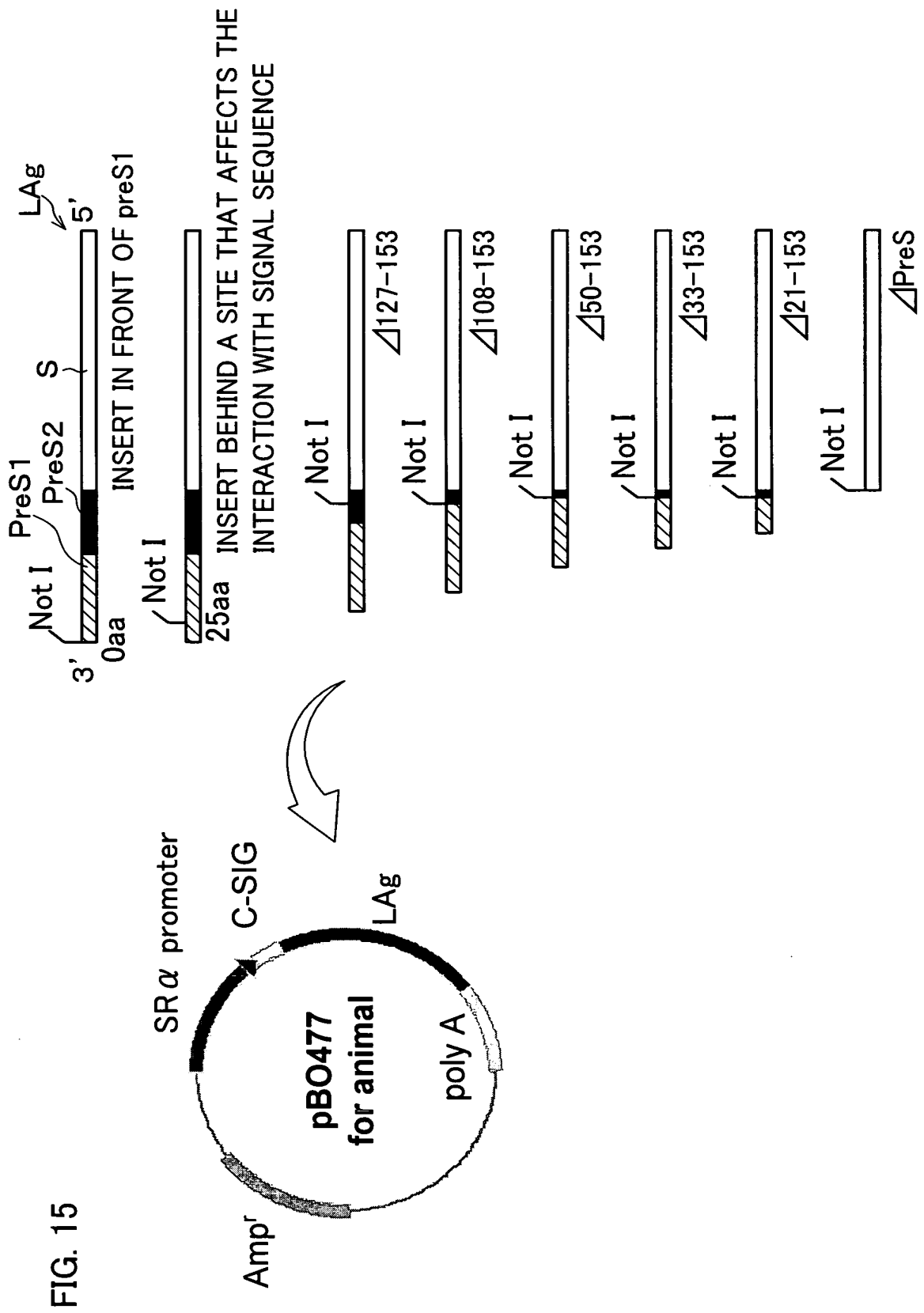


FIG. 15

FIG. 16 (a) SUPERNATANT (× 2 DILUTED)

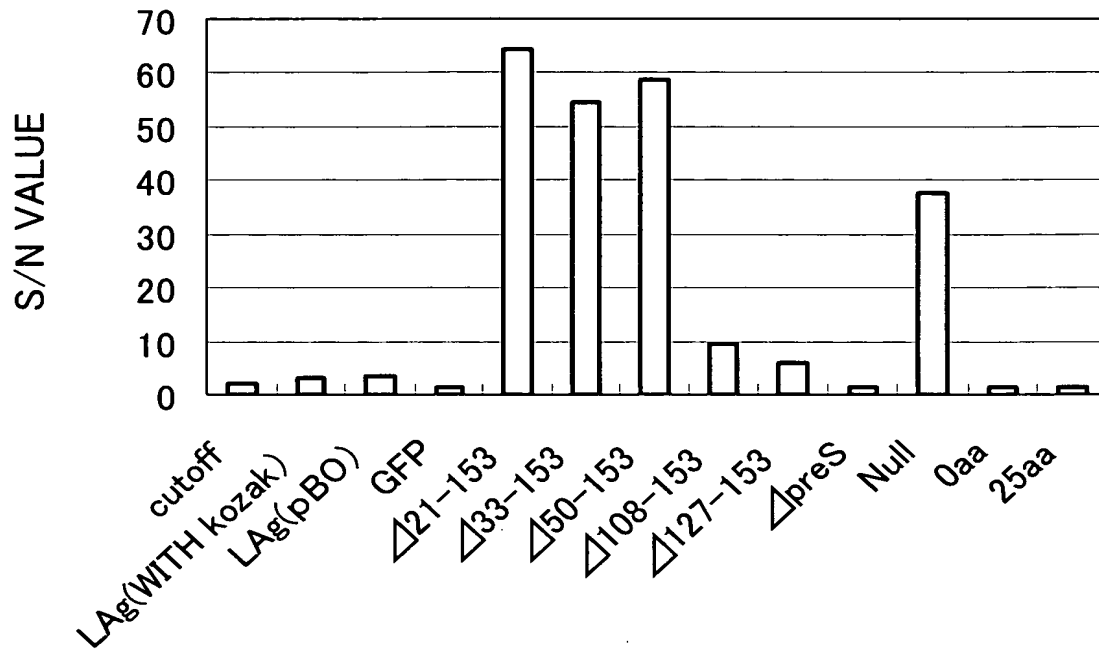


FIG. 16 (b) CELLS (× 200 DILUTED)

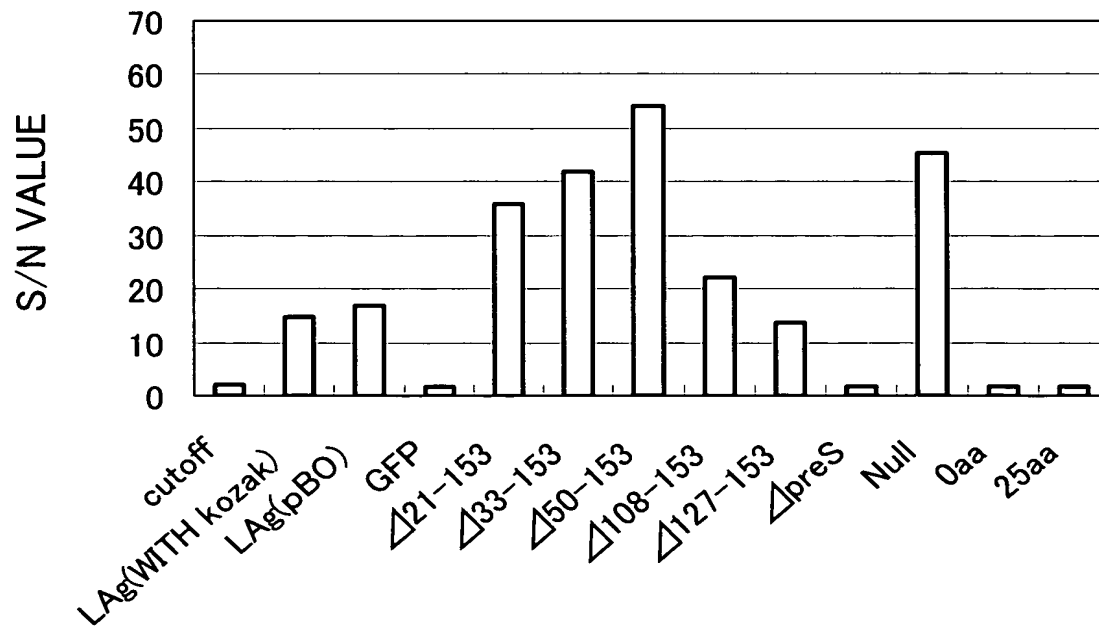


FIG. 17

plasmid	SUPERNATANT (× 2 DILUTED)			CELLS (× 200 DILUTED)		
	S/N	RATE	DECISION	S/N	RATE	DECISION
LA _g (WITH kozak)	3.13	26.6	R	14.74	115.0	R
LA _g (pBO)	3.68	31.3	R	17.00	132.6	R
GFP	1.41	12.0		1.69	13.2	
Δ ₂₁₋₁₅₃	64.54	548.6	R	35.79	279.2	R
Δ ₃₃₋₁₅₃	54.39	462.3	R	42.03	327.8	R
Δ ₅₀₋₁₅₃	58.67	498.7	R	54.19	422.7	R
Δ ₁₀₈₋₁₅₃	9.58	81.4	R	22.32	174.1	R
Δ ₁₂₇₋₁₅₃	5.93	50.4	R	13.73	107.1	R
Δ _{preS}	1.52	12.9		1.59	12.4	
Null	37.73	320.7	R	45.41	354.2	R
0aa	1.42	12.1		1.65	12.9	
25aa	1.46	12.4		1.6	12.5	

FIG. 18

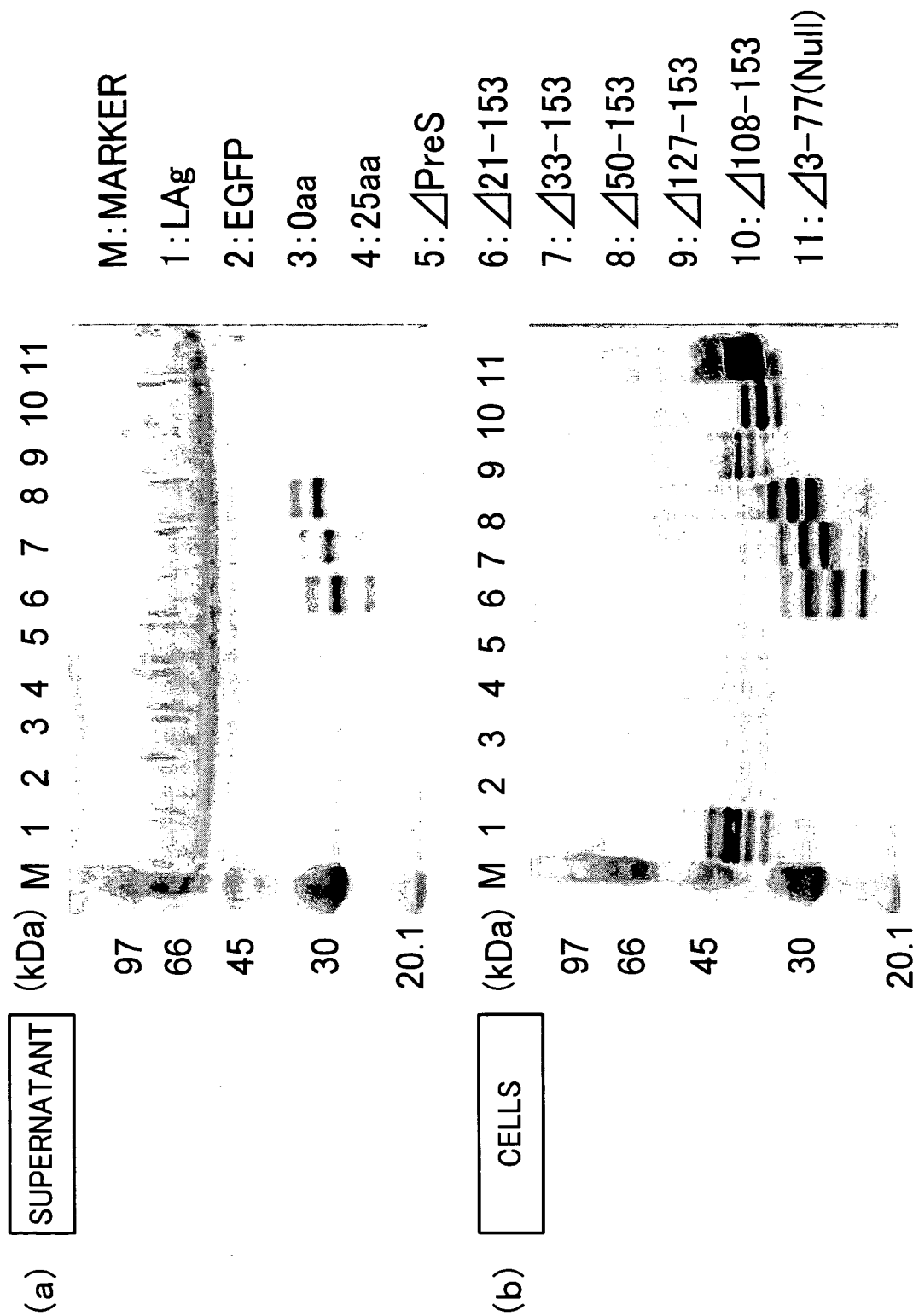


FIG. 19

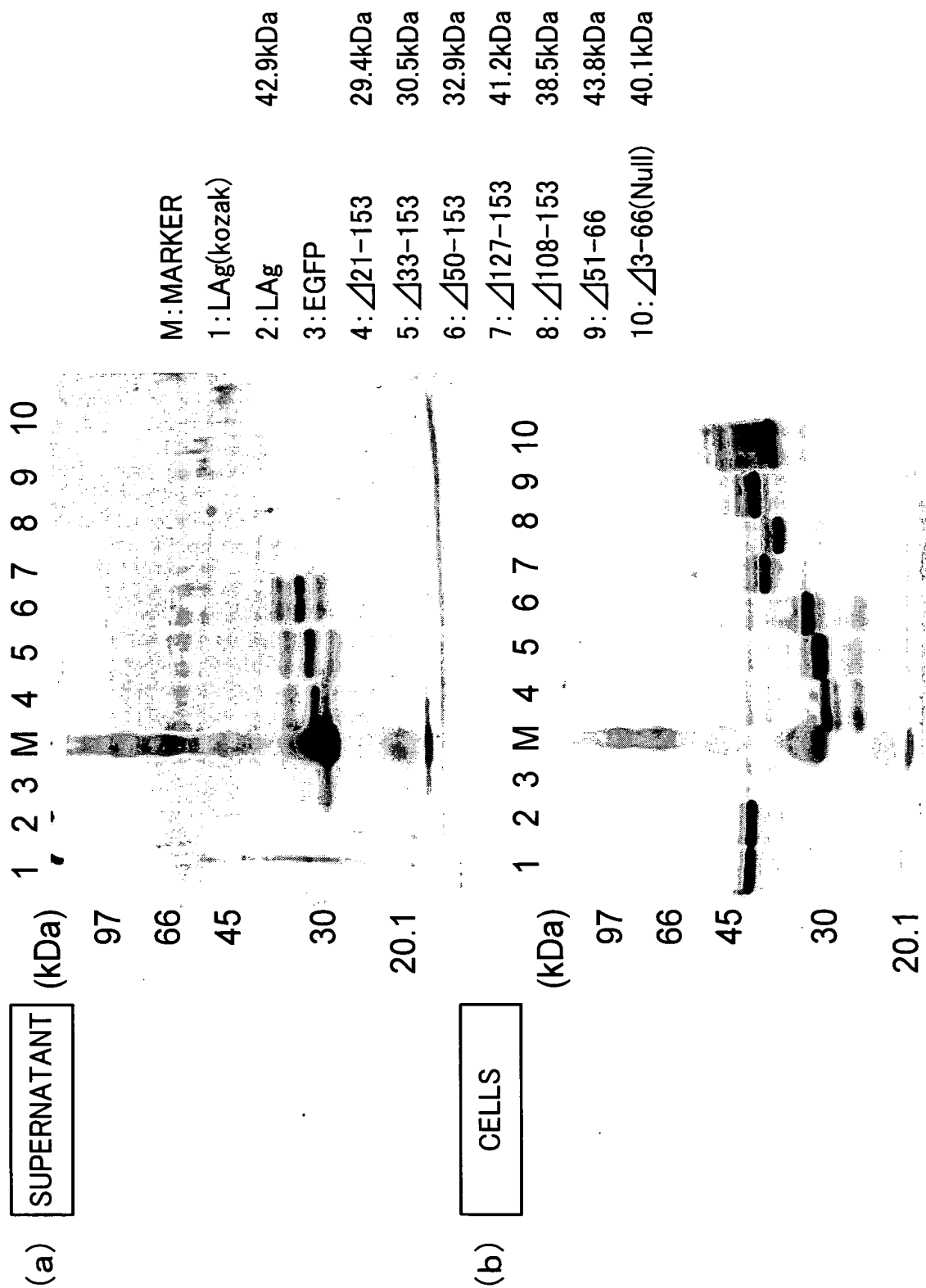
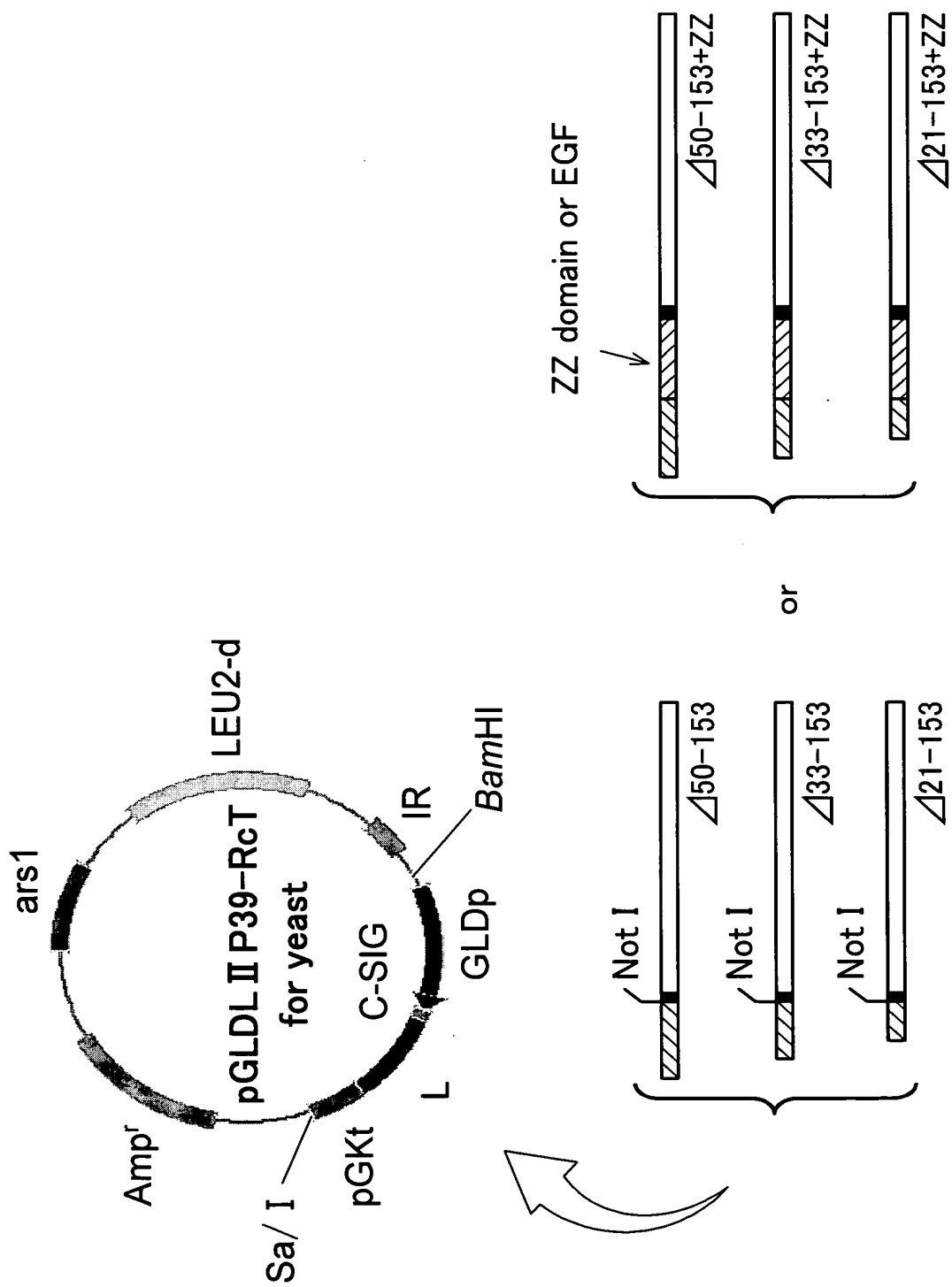


FIG. 20



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FIG. 21

STRAIN	S/N
LA _g	233
△21-153	255.03
△33-153	189.22
△50-153	318.45
△21-153+ZZ	...
△33-153+ZZ	280.18
△50-153+ZZ	304

FIG. 22

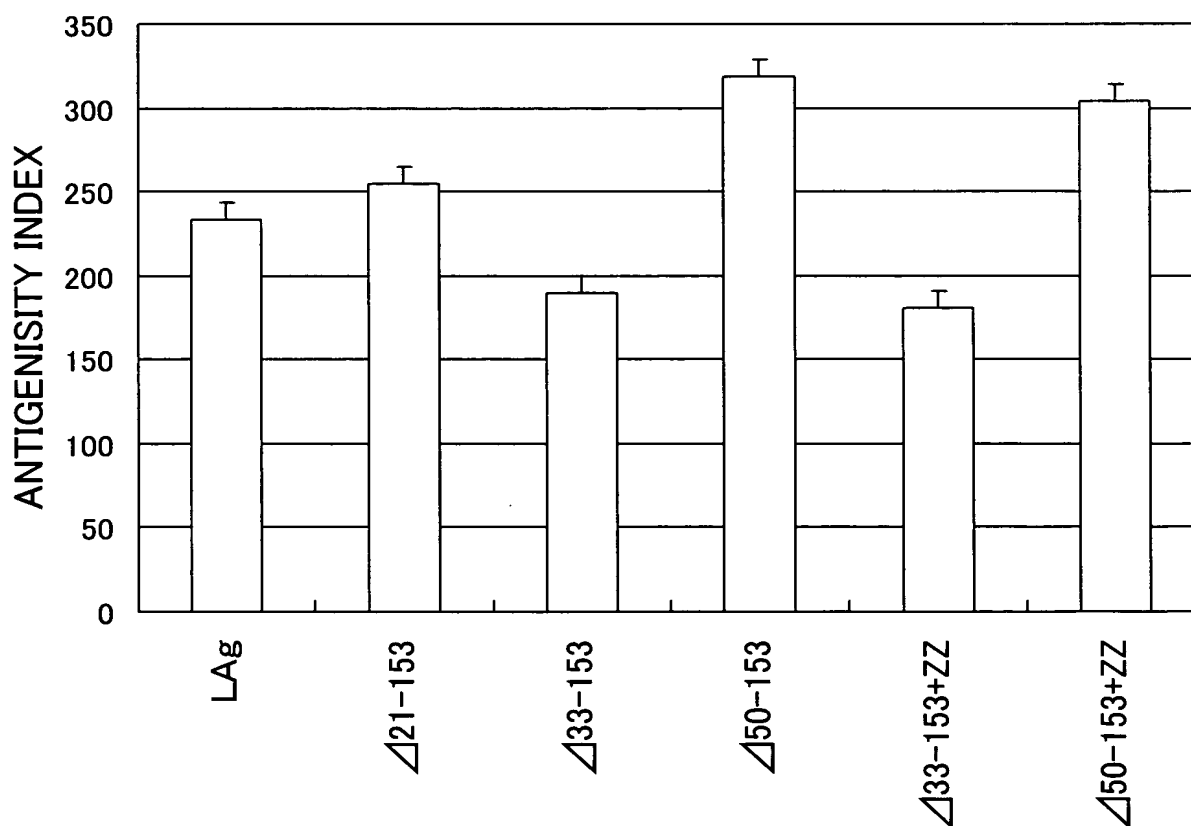


FIG. 23

PROTEINS ATTACKING CYTOPLASMIC RNA SUCH AS RNase	Pancreatic type Rnases from vertebrates
	RNase 1 or Bovine RNase A
	Eosinophil derived neurotoxin
	Eosinophil cationic protein
	Liver RNase (RNase 4)
	Angiogenin
	Bovine seminal RNase
	Frog Rnases (Onconase etc.)
PROTEINS OBSTRUCTING MEMBRANE TRANSPORT	Streptolysin(<i>Streptococcus pyogenes</i>)
	Cholesterol binding toxins (<i>Streptococcus</i> . <i>Bacillus</i> . <i>Clostridium</i> . <i>Listeria</i>)
	alpha-Toxin (<i>Staphylococcus aureus</i>)
	Delta-Toxin (<i>Staphylococcus aureus</i>) and melittin (<i>Apis mellifera</i>)
	Aerolysin (<i>Aeromonas hydrophila</i>)
	<i>Escherichia coli</i> hemolysin
PROTEINS OBSTRUCTING SIGNAL TRANSDUCTION	Cholera toxin (<i>Vibrio cholerae</i>)
	Heat-labile enterotoxins (<i>Escherichia Coli</i> LD
	Pertussis toxin (<i>Bordetella pertussis</i>)
	Exoenzyme C3 (<i>Clostridium botulinum</i>)
	Adenylate cyclase toxin (<i>Bordetella</i> sp.)
	Anthrax edema factor (<i>Bacillus anthracis</i>)
PROTEINS OBSTRUCTING PROTEIN SYNTHESIS	Diphtheria toxin (<i>Corynebacterium diphtheriae</i>)
	<i>Pseudomonas aeruginosa</i> exotoxin A
	Shiga toxins (<i>Shigella dysenteriae</i> serotype I, <i>Escherichia Coli</i>)
	Ricin (<i>Ricinus communis</i>)
	Ribosome-inactivating proteins
	alpha-Sarcin and related toxins (<i>Aspergillus</i>)
PROTEINS DISTURBING CYTOSKELTON	C2 toxin (<i>Clostridium botulinum</i> type C and D)
	Cytotoxic necrotizing factors (<i>Escherichia coli</i>)
	Enterotoxin A and cytotoxin B (<i>Clostridium</i> <i>difficile</i>)
	ActA (<i>Listeria monocytogenes</i>)
	IcsA (<i>Shigella flexneri</i>)
	Zonula occludens toxin (<i>Vibrio cholerae</i>)

FIG. 24

PROTEINS SUPPRESSING IMMUNITY OR INFLAMMATORY REACTION	Pyrogenic exotoxins (superantigens) (Staphylococcus aureus and Streptococcus pyogenes)
	Anthrax lethal toxin (Bacillus anthracis)
	Leukocidins and gamma lysins (Staphylococcus sp.)
PROTEINS DISTURBING MEMBRANE TRANSPORT	Tetanus neurotoxin (Clostridium tetani)
	VAMP-specific botulinum neurotoxins
	Botulinum neurotoxins type A and E (Clostridium botulinum)
	Botulinum neurotoxin type C (Clostridium botulinum)
	Vacuolating cytotoxin (Helicobacter pylori)
Na CHANNEL DISTURBING PROTEINS	alpha-Scorpion toxins
	beta-Scorpion toxins
	Excitatory insect selective neurotoxins from scorpion venoms
	Depressant insect selective neurotoxins from scorpion venoms
	mu-Conotoxins (Conus geographus)
	mu-Agatoxins (Agelenopsis aperta)
	Anthopleurin-A, -B, and -C (anemone toxin)
	Anemone toxins (type II)
	Calitoxins
K CHANNEL DISTURBING PROTEINS	Kaliotoxin
	Scyllatoxin (Leiurus quinquestriatus hebraeus)
	Apamin (honey bee Apis mellifera)
	MCD peptide (honey bee Apis mellifera)
	Charybdotoxin and iberiotoxin (Leiurus quinquestriatus var. hebraeus and Buthus tamulus)
	Margatoxin, noxiustoxin, and kaliotoxin (Centruroides margaritatus, Centruroides noxius, Androctonus mauretanicus)
	Dendrotoxins (Dendroaspis species)
	Sea anemone potassium channel toxins

FIG. 25

Ca CHANNEL DISTURBING PROTEINS	Omega-Conotoxins (<i>Conus</i> spp.)
	Omega-Agatoxins (<i>Agelenopsis aperta</i>)
	Omega-Grammotoxin SIA (<i>Grammostola spatulata</i> Chilean pink tarantula)
	Hololena toxin (<i>Hololena curta</i>)
	PLTXII (<i>Plectreurys tristes</i>)
	Calciseptine (<i>Dendroaspis polylepis</i>)
	Calcicludeine (<i>Dendroaspis angusticeps</i>)
	beta-Leptinotarsin-h
	Taicatoxin (<i>Oxyuranus scutellatus scutellatus</i>)
ACETYLCHOLINE RECEPTOR DISTURBING PROTEINS	alpha-Bungarotoxin (<i>Bungarus multicinctus</i>)
	alpha-Cobratoxin (<i>Naja kaouthia</i>)
	Erabutoxins (<i>Laticauda semifasciata</i>)
	Toxin alpha (' <i>Naja nigricollis</i> ')
	kappa-Bungarotoxin (<i>Bungarus multicinctus</i>)
	alpha-Conotoxins (<i>Conus</i> spp.)
	Snake toxins against muscarinic acetylcholine receptors
	Muscarinic toxin-1~5, -7, m1-toxin from green mamba (<i>Dendroaspis angusticeps</i>)
	Muscarinic toxin-alpha, -beta from black mamba (<i>Dendroaspis polylepis</i>)
RYANODINE RECEPTOR Ca ²⁺ CHANNEL DISTURBING PROTEINS	Helothermine (<i>Heloderma horridum horridum</i>)

FIG. 26

PRESYNAPTIC DISTURBING PROTEINS	PRESYNAPTIC DISTURBING PROTEINS
	Rattlesnake venom neurotoxins: crotoxin-related proteins
	Ammodytotoxins (<i>Vipera ammodytes ammodytes</i>)
	Notexins (<i>Notechis scutatus scutatus</i>)
	Textilotoxin (<i>Pseudonaja textilis textilis</i>)
	Tai poxin
	alpha-Latrotoxin (black widow spider)
	alpha-Latroinsectotoxin (<i>Latrodectus mactans tred ecimguttatus</i>)
	Pardaxin (<i>Pardachirus marmoratus</i>)
	Palytoxin (Corals of the spp. <i>Palythoa</i>)
GLUTAMIC ACID RECEPTOR DISTURBING PROTEINS	Equinatoxins (<i>Actinia equina</i> L., sea anemone)
	Conantokins (<i>Conus</i> spp.)

FIG. 27

		TIME AFTER GCV ADMINISTRATION (DAYS)						
		0	1	3	6	9	1 2	1 5
TUMOR SIZE (%)	A 4 3 1	100	102	97	94	85	72	66
	W i D r	100	105	107	111	118	125	133

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